IBC Meeting Minutes
6/11/2026 12:05 PM
CMRI Conference Room 31 & Teams

MEETING TIME RECORDS

Meeting start time: 12:05
Meeting end time: 13:30

VOTING MEMBER ATTENDANCE

Name of IBC Member	Committee Role	Late Arrival Time	Early Departure Time
Daniel Heruth	Chair, Animal Expert, Lab Rep		
Paul Ramlow	BSO, Animal Expert, Lab Rep		
Vivekanand Yadav	Alternate Lab Rep (Srivastava)		
Judy Dilts	Local Non-Affiliated		
Tamie Crutchfield	Occupational Health		

VOTING MEMBERS ABSENT

Tarak Srivastava	Lab Rep		
James Slaughter	Local Non-Affiliated		
Todd Bradley	Co-Chair, Lab Rep		

NON-VOTING MEMBERS PRESENT

Foster St. Claire	AV & Ex-Officio		

ADMINISTRATIVE STAFF

Saskia Miller	Director of the ORI		
Daniel Crabtree	Sr. IBC Manager		
Kelleigh Pearson	Administrative Project Coordinator		

QUORUM INFORMATION

Number of SAFETY members on the roster: 7

GUEST NAMES

Debarpan Dhar, Taeju Park

Conflict of Interest (COI): Chair reminded the Committee to declare any COI.

Previous Meeting minutes approved: Yes
A motion to approve was made and seconded. The motion passed with 5 votes in favor, 2 absent.

BIOSAFETY COMMITTEE UPDATES

1. **Biological Safety Officer (BSO) Updates**
   a. The BSO reported a laboratory [redacted] inciden [redacted] Mitigation measures and additional researcher education are being reviewed. The Occupational Health Director provided an overview of the post-incident response and follow-up actions. The Committee had no questions.
   b. The BSO also reported that construction of the 4th floor will begin, and blueprints will be shared with the IBC at the July meeting for review. The Chair added, the build-out of the 8th and 9th floors is scheduled to begin on July 1, 2027, with plans to be presented to the IBC closer to that date.
2. **Office of Research Integrity (ORI) Updates**
   a. Debarpan Dhar was introduced as a guest observing IBC operations.
   b. Research laboratory building layouts, organized by PI, are available in the IBC Administrative folder in Microsoft Teams in the Maps folder.

REVIEW OF SUBMISSIONS

De Novo Review

3. **Review of IBC00117**

Title:	Determination of host controls of antibody responses
Investigator:	Todd Bradley
Submission ID	IBC00117
Attempt to Express Foreign Genes:	None, only human cell culture.
Protein Produced:	ICAM-1, ICAM-5, IL32, RAB11, Human monoclonal antibodies, Enterovirus capsid proteins.
Agent Characteristics:	Recombinant Nucleic Acid Work: 1. We will perform PCR amplification and cloning of human genes into DNA expression vectors or lentiviral particles for gene overexpression and knockdown in cell culture. 2. We will also use immunoglobulin genes to transfect the heavy and light chain genes into 293 cells in order to express

	soluble antibody to be purified from the cell culture supernatant. 3. We will generate and use lipid nanoparticles (LNPs) containing mRNA in both cell culture and in mouse experiments to transiently transfect cells. Vectors: Lentivirus pLenti-GIII-CMV-C-term-HA pLenti-U6-sgRNA-SFFV-Cas9-2A-Puro All mRNAs for LNP experiments will be generated by Genscript. mRNAs will be delivered using lipid nanoparticles (not viral).
Agent Containment:	Biological Containment Levels: • Enterovirus: BSL-2 • Induced Excitatory Neurons: BSL-2 • Mouse Tissue: BSL-1 • Human Derived Blood and Blood Types: BSL-2 • Synovial Fluid: BSL-2 • SH-SY5Y: BSL-2 • THP-1: BSL-2 • Peripheral Blood Mononuclear Cells (PBMC): BSL-2 • Human induced pluripotent stem cells: BSL-2 • K-562: BSL-2 • NK-92: BSL-2 • A-549: BSL-2 • HL-60: BSL-2 • HEK-293: BSL-2 • RAJI: BSL-2 • Influenza virus type A (Orthomyxoviruses): BSL-2 • Lentivirus (specify generation): BSL-2 • Respiratory syncytial virus (Paramyxoviruses): BSL-2 • Influenza virus type B (Orthomyxoviruses): BSL-2 • Human immunodeficiency virus (HIV) types 2 (Retroviruses): BSL-2 • Metapneumovirus: BSL-2 • Microglia: BSL-2 • Forebrain organoids: BSL-2
Applicable NIH Guidelines:	• Section III-D-1 • Section III-D

Determination: Modifications Required

Required modifications:

- Remove reference to UV light as a decontamination method.
- Revise disinfectant language:

- Clarify 10% bleach is for decontamination, not routine surface wiping
- Add use of 70% ethanol (or appropriate disinfectant) for surface cleaning
- Remove statement indicating solids can be treated with 10% bleach and revise accordingly.

Comments:

The BSO provided an overview of the protocol to the Committee.

The BSO noted that UV light is not a reliable method for decontamination.

The BSO clarified that 10% bleach should be used for decontamination purposes, but is not recommended for routine surface wiping, and that 70% ethanol (EtOH) or other appropriate disinfectants are better suited for that purpose.

The BSO further stated that solid materials should not be treated with 10% bleach and requested that this statement be removed and revised in the protocol.

Following discussion, a motion was made to require modifications to secure approval. The motion was seconded and passed with a vote of 5 in favor and 2 absent.

Supporting documents:

None

Votes:

For:	5
Against:	0
Recused:	0
Absent:	2
Abstained:	0

De Novo Review

4. **Review of IBC00120**

Title:	Preparation, genotyping and expression of nucleic acids from various resources.
Investigator:	Erin Boone
Submission ID	IBC00120
Attempt to Express Foreign Genes:	none
Protein Produced:	none
Agent Characteristics:	Recombinant Nucleic Acid Work: Extracted human genomic DNA or cDNA will be PCR amplified for genotyping, expression and/or sequencing. In some cases PCR amplicons will subsequently be cloned into plasmid vectors for more specific allele or splice variant sequencing. Most genotyping is

	done with commercially available assays. If none are available, primers will be designed in-house. The plasmid vectors are all commercially available. Vectors: pBR or M13 derived plasmid vectors will be used, i.e., pcDNA 3.1, pCR4-TOPO, pCR-XL-TOPO, pBS and other related commercially available vectors
Agent Containment:	Biological Containment Levels: • Saliva: BSL-2 • Other: BSL-2 • Urine: BSL-2 • Human Derived Blood and Blood Types: BSL-2 • Escherichia coli K12 or derivative: BSL-1
Applicable NIH Guidelines:	• Section III-F • Section III-F-8

Determination: Modifications Required

Required modifications:

- Clarify TB risk from lung tissue; include or rule out and add bloodborne pathogen risk language as appropriate.
- Clarify use of pcDNA3.1: either explain its use with mammalian cells or remove it.
- Specify any cell lines used; if none, state that work is limited to bacterial expression.
- Ensure all sections are consistent (materials, methods, and risks align).

Comments:

The BSO provided an overview of the protocol to the committee.

In response to a question regarding the necessity of surgical masks, the BSO explained the rationale and mitigation measures in place that support not requiring mask use. The Committee accepted this explanation.

A committee member inquired whether there is a specific reason to suspect that the lung tissue involved could contain Mycobacterium tuberculosis (TB). The member recommended broadening the risk description to include bloodborne pathogens and clarifying the potential risk of TB.

A committee member noted that pcDNA3.1, a mammalian expression vector, is listed in the protocol; however, no mammalian cell lines are identified. The member requested clarification or removal of this vector from the protocol.

Another member observed that the protocol references expression in cell lines, but no specific cell lines (e.g., human or mouse) are described. The member recommended clarifying whether expression work is limited to bacterial systems and, if so, specifying this in the protocol.

Following discussion, a motion was made to require modifications to secure approval. The motion was seconded and passed with a vote of 5 in favor and 2 absent.

Supporting documents:
None

Votes:

For:	5
Against:	0
Recused:	0
Absent:	2
Abstained:	0

De Novo Review

5. **Review of IBC00119**

Title:	Regulation of Human ADMET
Investigator:	Erin Boone
Submission ID	IBC00119
Attempt to Express Foreign Genes:	All genes in the described experiments will be transiently introduced in to tissue culture cell lines.
Protein Produced:	Transcriptional regulators, reporter genes (i.e. Firefly and Renilla luciferases) and dCas9 fusion proteins will be transiently expressed in human cells into which plasmid constructs have been introduced.
Agent Characteristics:	Recombinant Nucleic Acid Work: Recombinant/synthetic nucleic acid work employs cultured cell models to investigate the molecular mechanisms of developmental regulation and interindividual variability of human ADMET pathways. This work involves the use of the following: 1) promoter-reporter gene assays in transiently transfected human cell lines via lipid-mediated introduction of reporter plasmids and/or transcriptional activators 2) transient manipulation of epigenetic modifications (such as DNA methylation and histone modifications) using the Casilio system introduce via lipid-mediated methods. Vectors: Plasmid vectors to be used: pcDNA3.1, pGL3, pRL, pCR4-TOPO, pCR8, pEF1, pCR-XL-TOPO, pSG5, pBS, pENTR5'/CMVp, pENTR5'/EF1ap, pDPNR221, and related commercially available vectors

Agent Containment:	Biological Containment Levels: • Human Derived Blood and Blood Types: BSL-2 • Urine: BSL-1 • Digestive Tissue: BSL-2 • Mucus Membrane Tissue: BSL-2 • CACO-2: BSL-2 • A-549: BSL-2 • COS-7: BSL-2 • Human Primary Liver Epithelial Cells: BSL-2 • HEP-3B: BSL-2 • HEP-G2: BSL-2 • HEK-293T: BSL-2 • Escherichia coli K12 or derivative: BSL-1
Applicable NIH Guidelines:	• Section III-F • Section III-F-1 • Section III-F-8

Determination: Modifications Required

Required modifications:

- Add more detail to the research summary (explain ADMET and expand bullet 2).
- Remove the PAPR reference from the protocol.
- Update risk section to general bloodborne pathogen language and clarify TB risk for lung samples.
- Add details on shipping/transport procedures, including containers and CMRI protocol reference.
- List only vectors actually used in the lab (include related ones only if relevant).
- Provide the maximum or range of sample numbers for each tissue type in the lab.

Comments:

The BSO provided an overview of the protocol to the Committee.

A member noted that the research summary requires additional detail, specifically:

-Clarification of the importance of ADMET in bullet one

-Additional explanation for bullet two

A member questioned the inclusion of a PAPR. The BSO reviewed the section in question and agreed it should not be listed. The Committee requested removal of the PAPR reference.

A member recommended revising the risk description to reflect a general bloodborne pathogen risk rather than identifying specific pathogens. The member also requested clarification regarding the potential for increased TB risk in lung specimens.

A member requested additional detail on shipping and transport procedures, including containers used and reference to the CMRI shipping and transport protocol.

A member stated that only vectors actively used in the laboratory should be listed; related vectors may be included if applicable.

A member requested that the PI provide the maximum or range of quantities for each tissue type currently maintained in the lab.

Following discussion, a motion was made to require modifications to secure approval. The motion was seconded and passed with a vote of 5 in favor and 2 absent.

Supporting documents:
None

Votes:

For:	5
Against:	0
Recused:	0
Absent:	2
Abstained:	0

De Novo Review

6. **Review of IBC00121**

Title:	Normal and Neoplastic Growth
Investigator:	Taeju Park
Submission ID	IBC00121
Attempt to Express Foreign Genes:	- We transfect mammalian cells with recombinant or synthetic DNAs or RNAs to control protein expression and study signal transduction pathways at the cellular level. - We have generated various genetically engineered mouse models at previous institutions, and those mouse strains are currently preserved as frozen embryos and sperm in liquid nitrogen freezers. - We have generated model strains and maintained the mouse strains.
Protein Produced:	Mouse Crk and CrkL proteins were purified from bacterial cells. Synthetic proteins were purified from bacterial cells.

Agent Characteristics:	Recombinant Nucleic Acid Work: We generate various synthetic nucleic acids by amplifying cDNAs using PCR and cloning them into mammalian expression vectors. Plasmid DNAs are amplified in bacterial cells, and mRNAs are synthesized in vitro. Mammalian cells are transfected with DNAs or synthetic mRNAs to express proteins. We also transfect mammalian cells with commercially available small interfering RNA (siRNA) to suppress protein expression. For transfections, we use calcium phosphate, Lipofectamine, FuGENE 6, X-tremeGENE 9, and electroporation methods. Vectors (non-viral): pGEM-T Easy, TOPO, pDS56, pGEX, pCMV, and pcDNA3 and other mammalian expression vectors derived from pcDNA3 Bacterial hosts: - Escherichia coli (XL1-Blue and XL10-Gold) for introduction of site-specific point or deletion mutations - Escherichia coli (DH5 alpha, One Shot Top10, and Mach1) for cloning and amplification of plasmid DNAs - Escherichia coli (BL21(DE3)) for protein synthesis induction using IPTG and protein purification Mammalian cells for protein expression: - Commercially available human cell lines - Commercially available mouse cell lines - Commercially available human glioblastoma and glioma cell lines - Glioma cell lines obtained from an academic research laboratory or a multi-institutional, patient-driven research network - Lab-established mouse embryonic fibroblasts (MEFs) - Lab-established human and mouse brain tumor cell lines Vectors: Vectors (non-viral): pGEM-T Easy, TOPO, pDS56, pGEX, pCMV, pET-30a(+), pcDNA3 and other mammalian expression vectors derived from pcDNA3 - Lentiviral vector: pLVX and Lenti-X HTX packaging system (Takara Bio) - Retroviral vector: pBABE-Puro and Platinum retroviral packaging cell lines (Cell Biolabs, Inc) - Retroviral vector: pSuper.retro.neo (OligoEngine) and Phoenix helper-free retrovirus producer lines (ATCC) - Lentiviral vector: pLV[shRNA] and pLV[Exp] (both from VectorBuilder)

Agent Containment:	Biological Containment Levels: • 7316-440: BSL-2 • 7316-161: BSL-2 • 7316-4917: BSL-2 • 7316-2176: BSL-2 • Mouse Tissue: BSL-2 • Cerebrospinal Fluid: BSL-2 • Human Derived Blood and Blood Types: BSL-2 • Human Tissue: BSL-2 • HK-2: BSL-2 • CAKI-1: BSL-2 • U-87MG: BSL-2 • WT 9-12: BSL-2 • SF8628: BSL-2 • SU-DIPG XIII: BSL-2 • SU-DIPG VI: BSL-2 • SU-DIPG IV: BSL-2 • LN-229: BSL-2 • A-549: BSL-2 • Hs 683 (human glioma cell line): BSL-2 • 4T1 (mouse breast cancer cell line): BSL-2 • NIH3T3 (mouse fibroblast cell line): BSL-2 • U-118MG (human glioblastoma cell line): BSL-2 • HELA: BSL-2 • HEK-293T: BSL-2 • Escherichia coli (XL1-Blue and XL10-Gold): BSL-1 • Escherichia coli (DH5 alpha, One Shot Top10, and Mach1): BSL-1 • Escherichia coli (BL21(DE3)): BSL-1 • pLV lentivirus: BSL-2 • pBABE retrovirus: BSL-2 • pLVX lentivirus: BSL-2 • pSUPER retrovirus: BSL-2
Applicable NIH Guidelines:	• Section III-D-1-a • Section III-E-3 • Section III-F-3 • Section III-D-4 • Section III-F • Section III-E • Section III-F-1 • Section III-D-1

Determination: Modifications Required

Required modifications:

- Remove language describing likelihood of exposure; focus only on consequences of exposure.

- Revise biosafety level for mouse biospecimens from BSL-2 to BSL-1.
- Update the short title to match the full title.
- Provide an estimated maximum number of samples for each tissue/specimen type (e.g., total blood samples stored).
- Clarify tissue quantities (e.g., mg amounts vs. cell suspensions) and revise accordingly.

Comments:
Dr. Park was introduced and provided an overview of his protocol and research. Following questions from the IBC, the Committee thanked Dr. Park for his presentation. Dr. Park exited the meeting at 12:30 PM.

The BSO then provided an overview of the protocol to the Committee.

A member requested that language describing the likelihood of exposure be removed and replaced with a focus on the consequences of exposure.

A member noted that the PI selected BSL-2 for mouse biospecimens, which are typically handled at BSL-1. Following discussion and review, the Committee agreed the appropriate level is BSL-1.

A member recommended that the short title match the full title, noting that the abbreviated version appears incomplete. The Committee agreed.

A member requested that the PI provide an estimated maximum number of samples for each tissue type maintained in the laboratory (e.g., total number of blood samples).

A member asked for clarification on how tissue quantities are reported (e.g., mg vs. cell suspensions) and requested that the protocol be revised accordingly.

Following discussion, a motion was made to require modifications to secure approval. The motion was seconded and passed with a vote of 5 in favor and 2 absent.

Supporting documents:
None

Votes:

For:	5
Against:	0
Recused:	0
Absent:	2
Abstained:	0

REVIEW OF OTHER AGENDA ITEMS

ACKNOWLEDGMENTS SECTION

The following items were approved outside of a convened IBC meeting. The Committee reviewed and acknowledged the following items during this meeting.

ID	Name	State	Specialist	Agenda Type
SAMEND202600023	Amendment for IBC00100	Approved	Daniel Crabtree	Admin Approval
SAMEND202600025	6-26 Amendment for IBC00099	Approved	Daniel Crabtree	Admin Approval
SAMEND202600027	Amendment for IBC00108	Approved	Elizabeth Short	Admin Approval
SAMEND202600028	Amendment for IBC00111	Approved	Elizabeth Short	Admin Approval
SAMEND202600029	Amendment for IBC00084 - Summer Scholar	Approved	Elizabeth Short	Admin Approval
SAMENDCR202600000002	Amendment/CR for IBC00114	Approved	Daniel Crabtree	Admin Approval